

## Chapter 3

### General

#### Active transport

Stasi, R., Neves, H. I. & Spira, B. (2019). Phosphate uptake by the phosphonate transport system PhnCDE. *BMC Microbiology* **19**(1), 79. <https://doi.org/10.1186/s12866-019-1445-3>

Vuppada, R. K., *et al.* (2018). Phosphate signaling through alternate conformations of the PstSCAB phosphate transporter. *BMC Microbiology* **18**(1), 8. <https://doi.org/10.1186/s12866-017-1126-z>

#### ATP-binding cassette (ABC) pathway

Delepelaire, P. (2019). Bacterial ABC transporters of iron containing compounds. *Research in Microbiology* **170**(8), 345-357. <https://doi.org/10.1016/j.resmic.2019.10.008>

Mächtel, R., *et al.* (2019). An integrated transport mechanism of the maltose ABC importer. *Research in Microbiology* **170**(8), 321-337. <https://doi.org/10.1016/j.resmic.2019.09.004>

#### Tripartite ATP-independent periplasmic (TRAP) transporters

Schäfer, L., *et al.* (2019). A tripartite tricarboxylate transporter (MIM\_c39170–MIM\_c39210) of *Advenella mimigardefordensis* DPN7T is involved in citrate uptake. *International Microbiology* **22**(4), 461-470. <https://doi.org/10.1007/s10123-019-00073-5>

## **Group translocation**

## **Iron uptake and siderophores**

Delepelaire, P. (2019). Bacterial ABC transporters of iron containing compounds. *Research in Microbiology* **170**(8), 345-357. <https://doi.org/10.1016/j.resmic.2019.10.008>

Smith, A. T., *et al.* (2019). The FeoC [4Fe–4S] cluster is redox-active and rapidly oxygen-sensitive. *Biochemistry* **58**(49), 4935-4949. <https://doi.org/10.1021/acs.biochem.9b00745>

## **TonB-dependent active transport across the outer membrane in Gram-negative bacteria**

## **Multidrug efflux pump**

Orelle, C., Mathieu, K. & Jault, J.-M. (2019). Multidrug ABC transporters in bacteria.

*Research in Microbiology* **170**(8), 381-391.

<https://doi.org/10.1016/j.resmic.2019.06.001>

Severi, E. & Thomas, G. H. (2019). Antibiotic export: transporters involved in the final step of natural product production. *Microbiology* **165**(8), 805-818.

<https://doi.org/10.1099/mic.0.000794>

## **Protein translocation**

### **General secretion pathway (GSP)**

### **Twin-arginine translocation (TAT) pathway**

Dolata, K. M., *et al.* (2019). Far-reaching cellular consequences of *tat* deletion in

*Escherichia coli* revealed by comprehensive proteome analyses. *Microbiological*

*Research* **218**, 97-107. <https://doi.org/10.1016/j.micres.2018.10.008>

Ryu, J., *et al.* (2019). Tat-dependent heterologous secretion of recombinant tyrosinase by

*Pseudomonas fluorescens* is aided by a translationally fused caddie protein. *Applied and Environmental Microbiology* **85**(20), e01350-01319.

<https://aem.asm.org/content/aem/85/20/e01350-19.full.pdf>

Valverde, J. R., Gullón, S. & Mellado, R. P. (2018). Modelling the metabolism of protein secretion through the Tat route in *Streptomyces lividans*. *BMC Microbiology* **18**(1), 59. <https://doi.org/10.1186/s12866-018-1199-3>

### **Protein translocation through the ABC pathway**

Beis, K. & Rebuffat, S. (2019). Multifaceted ABC transporters associated to microcin and bacteriocin export. *Research in Microbiology* **170**(8), 399-406.  
<https://doi.org/10.1016/j.resmic.2019.07.002>

Geise, H., *et al.* (2019). A potential late stage intermediate of twin-arginine dependent protein translocation in *Escherichia coli*. *Frontiers in Microbiology* **10**, 1482.  
<https://www.frontiersin.org/article/10.3389/fmicb.2019.01482>

Heinkel, F. *et al.* (2019). Phase separation and clustering of an ABC transporter in *Mycobacterium tuberculosis*. *Proceedings of the National Academy of Sciences of the USA* **116**(33), 16326-16331.  
<https://www.pnas.org/content/pnas/116/33/16326.full.pdf>

Slamti, L. & Lereclus, D. (2019). The oligopeptide ABC-importers are essential communication channels in Gram-positive bacteria. *Research in Microbiology* **170**(8), 338-344. <https://doi.org/10.1016/j.resmic.2019.07.004>

van Veen, H. W., *et al.* (2019). Energy coupling in ABC exporters. *Research in Microbiology* **170**(8), 392-398. <https://doi.org/10.1016/j.resmic.2019.08.003>

## Protein translocation through the cell wall in Gram-positive bacteria

## Protein translocation in Gram-negative bacteria

Allsopp, L. P., Bernal, P., Nolan, L. M. & Filloux, A. (in press). Causalities of war: The connection between T6SS and microbiota. *Cellular Microbiology*.

<https://onlinelibrary.wiley.com/doi/abs/10.1111/cmi.13153>

Bayer-Santos, E., Ceseti, L. d. M., Farah, C. S. & Alvarez-Martinez, C. E. (2019).

Distribution, function and regulation of type 6 secretion systems of *Xanthomonadales*. *Frontiers in Microbiology* **10**, 1635.

<https://www.frontiersin.org/article/10.3389/fmicb.2019.01635>

Butan, C., *et al.* (2019). High-resolution view of the type III secretion export apparatus in situ reveals membrane remodeling and a secretion pathway. *Proceedings of the National Academy of Sciences of the USA* **116**(49), 24786-24795.

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Chen, C., Yang, X. & Shen, X. (2019). Confirmed and potential roles of bacterial T6SSs in the intestinal ecosystem. *Frontiers in Microbiology* **10**, 1484.

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Kirchweiger, P., *et al.* (2019). Structural and functional characterization of SiiA, an auxiliary protein from the SPI4-encoded type 1 secretion system from *Salmonella enterica*. *Molecular Microbiology* **112**(5), 1403-1422.

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Liang, X., *et al* (2019). An onboard checking mechanism ensures effector delivery of the type VI secretion system in *Vibrio cholerae*. *Proceedings of the National Academy of Sciences of the USA* **116**(46): 23292-23298.

<https://www.pnas.org/content/pnas/116/46/23292.full.pdf>

Liaw, J., *et al.* (2019). The *Campylobacter jejuni* type VI secretion system enhances the oxidative stress response and host colonization. *Frontiers in Microbiology* **10**, 2864.

<https://www.frontiersin.org/article/10.3389/fmicb.2019.02864>

Palmer, A. D., Kim, K. & Slauch, J. M. (2019). PhoP-mediated repression of the SPI1 type 3 secretion System in *Salmonella enterica* serovar Typhimurium. *Journal of Bacteriology* **201**(16), e00264-00219. <https://jb.asm.org/content/jb/201/16/e00264-19.full.pdf>

Peñil-Celis, A. & Garcillán-Barcia, M. P. (2019). Crosstalk between type VI secretion system and mobile genetic elements. *Frontiers in Molecular Biosciences* **6**, 126.

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Sharp, J. S., Rietsch, A. & Dove, S. L. (2019). RNase E promotes expression of type III secretion system genes in *Pseudomonas aeruginosa*. *Journal of Bacteriology* **201**(22), e00336-00319. <https://jb.asm.org/content/jb/201/22/e00336-19.full.pdf>

Stietz, M. S., *et al.* (2019). Double tubular contractile structure of the type VI secretion system displays striking flexibility and elasticity. *Journal of Bacteriology* **202**(1), e00425-00419. <https://jb.asm.org/content/jb/202/1/e00425-19.full.pdf>

Xie, Y., Shao, X. & Deng, X. (2019). Regulation of type III secretion system in *Pseudomonas syringae*. *Environmental Microbiology* **21**(12), 4465-4477.

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## **Type VII secretion system**

Famelis, N., *et al.* (2019). Architecture of the mycobacterial type VII secretion system.

*Nature* **576**(7786), 321-325. <https://doi.org/10.1038/s41586-019-1633-1>

## **Export of polysaccharides and components of surface structures**

Hughes, G. W., *et al.* (2019). Evidence for phospholipid export from the bacterial inner membrane by the Mla ABC transport system. *Nature Microbiology* **4**(10), 1692-1705.

<https://doi.org/10.1038/s41564-019-0481-y>

Sperandeo, P., Martorana, A. M. & Polissi, A. (2019). The Lpt ABC transporter for lipopolysaccharide export to the cell surface. *Research in Microbiology* **170**(8), 366-373.

<https://doi.org/10.1016/j.resmic.2019.07.005>

## **Protein secretion in Archaea**

## **Metallochaperones**

Antoine, R., Rivera-Millot, A., Roy, G. & Jacob-Dubuisson, F. (2019). Relationships between copper-related proteomes and lifestyles in  $\beta$  proteobacteria. *Frontiers in Microbiology* **10**, 2217. <https://www.frontiersin.org/article/10.3389/fmicb.2019.02217>

Christenson, E. T., *et al.* (2019). The iron-regulated vacuolar *Legionella pneumophila* MavN protein is a transition-metal transporter. *Proceedings of the National Academy of Sciences of the USA* **116**(36), 17775-17785.